Removal of Pharmaceutical Residues by Ferrate(VI)

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Abstract

Background: Pharmaceuticals and their metabolites are inevitably emitted into the waters. The adverse environmental and human health effects of pharmaceutical residues in water could take place under a very low concentration range; from several μg/L to ng/L. These are challenges to the global water industries as there is no unit process specifically designed to remove these pollutants. An efficient technology is thus sought to treat these pollutants in water and waste water.

Methodology/Major Results: A novel chemical, ferrate, was assessed using a standard jar test procedure for the removal of pharmaceuticals. The analytical protocols of pharmaceuticals were standard solid phase extraction together with various instrumentation methods including LC-MS, HPLC-UV and UV/Vis spectroscopy. Ferrate can remove more than 80% of ciprofloxacin (CIP) at ferrate dose of 1 mg Fe/L and 30% of ibuprofen (IBU) at ferrate dose of 2 mg Fe/L. Removal of pharmaceuticals by ferrate was pH dependant and this was in coordinate to the chemical/physical properties of pharmaceuticals. Ferrate has shown higher capability in the degradation of CIP than IBU; this is because CIP has electron-rich organic moieties (EOM) which can be readily degraded by ferrate oxidation and IBU has electron-withdrawing groups which has slow reaction rate with ferrate. Promising performance of ferrate in the treatment of real waste water effluent at both pH 6 and 8 and dose range of 1–5 mg Fe/L was observed. Removal efficiency of ciprofloxacin was the highest among the target compounds (63%), followed by naproxen (43%). On the other hand, n-acetyl sulphamethoxazole was the hardest to be removed by ferrate (8% only).

Conclusions: Ferrate is a promising chemical to be used to treat pharmaceuticals in waste water. Adjusting operating conditions in terms of the properties of target pharmaceuticals can maximise the pharmaceutical removal efficiency.

Introduction

Pharmaceuticals such as antibiotics, anti-inflammatory drugs, β-blockers and X-ray contrast media are widely used in our daily life. These pharmaceuticals and their metabolites cannot be fully utilized by human beings or animals and are inevitably emitted into the waters by excretion [1–4] and/or through the discharge of industry effluents and hospitals waste waters [5,6]. Results of toxicology studies have revealed that some pharmaceuticals are suspected to have direct toxicity to certain aquatic organisms [7–9] and they could accumulate slowly, and finally lead to irreversible change on wildlife and human beings [10]. The adverse environmental and human health effects could take place under a very low concentration range; from several μg/L to ng/L. Due to this, pharmaceutical and personal care products (PPCPs) and endocrine disrupting chemicals (EDCs) are classified as emerging micro-pollutants which have been a significant issue of environmental and public health concern.

The presence of emerging micro pollutants and their potential toxicity are challenge to the global water industries as there is no unit process specifically designed to remove these pollutants; activated sludge and secondary sedimentation in most wastewater treatment works (WWTWs) seems to be inefficient to eliminate them [11–15]. Thus, a number of recent studies have been carried out to explore suitable technologies to treat pharmaceutical residuals from water and wastewater. Ozonation was found to be effective to remove pharmaceuticals in municipal WWTWs [16]. Nanofiltration (NF) and reverse osmosis (RO) membrane filtration have been applied at bench, pilot and full scale [17]. Activated carbon adsorption [18] has also been proved as an efficient process to remove pharmaceuticals; addition of 5 mg/L of powder activated carbon with a 4-h contact time removed 50% to >98% of the volatile PPCPs and 10% to >95% of the polar PPCPs [19].

Another promising technology which can address the concerns on pharmaceutical residues is ferrate (VI) (Fe(VI)). Ferrate (VI) exhibits many advantages because of its dual function as an oxidant and a coagulant [20–24] and oxidation of carbohydrates [31] and nitrogen-containing pollutants [32]. And research on the removal of pharmaceuticals and other micro pollutants by ferrate (VI) have been conducted recently (e.g., [33–38]). Moreover, ferrate (VI) is capable of removing more than 85% of various micro-pollutants containing electron-rich moieties (ERM) [39].

This paper aims to address the issue of the performance optimisation of pharmaceutical treatment by ferrate (VI). The dose of ferrate, test water pHs and the physical and chemical properties of pharmaceuticals are major factors to influence the overall treatment efficiency and these were studied in this work.
Materials and Reagents
Ciprofloxacin (98%, HPLC grade), ibuprofen (98%, GC grade), ibuprofen sodium salt (iBu-Na) and potassium ferrate (VI) (>90%) were purchased from Sigma-Aldrich; ciprofloxacin hydrochloride (CIP-HCl) was purchased from VWR; other chemicals and reagents used were obtained from Fisher Scientific. All chemicals and reagents were used without further purification. Deionised water used was generated by Elgacan B114 deioniser.

Ferrate Working Solution and Model Test Solution
The ferrate (VI) working solution (1 g/L) was prepared by the addition of solid K2FeO4 to 0.005 M Na2HPO4/0.001 M borate buffer solution at pH 9.0, the pH at which the ferrate solutions are most stable [40].

Both CIP and IBU stock solutions were prepared separately in deionised water with the concentration of 10 mg/L. Several types of model test solution samples (Table 1) were prepared by the dilution of stock solutions to 1 L with tap water, with the solution pH (6.8 – 7.3) unadjusted before dosing ferrate (VI).

Real Wastewater Effluents and Target Pharmaceuticals
The wastewater samples were taken from the second sedimentation effluent at Shieldhall WWTW, Glasgow, and the major properties of the samples were COD = 35 mg/L, turbidity = 2 NTU and pH was 7.37.

Two litres of raw samples were analysed for pharmaceutical concentrations, and the sample was then spiked with selected pharmaceuticals; each with concentration of 10 μg/L. The spiked pharmaceuticals are as follows, 1) X-ray contrast media: Iopamidol, Ammonium diatrizoate; 2) Antibiotics: Ciprofloxacin, Sulphamethoxazole, N-acetyl sulphamethoxazole, Erythromycin-H2O; 3) Non-steroidal anti-inflammatory drugs (NSAID): Naproxen, Ibuprofen; 4) β-blockers: Atenolol; 5) Antiepileptics: Carbamazepine; 7) Blood Lipid Lowering: Bezafibrate and 8) Local anaesthetic: Lidocaine.

Jar Test
A series of jar test experiments was carried out with a six-unit stirrer (Kemirafloculator 2000, Kemwater) under the following protocol: fast mixing for 1 min at 400 rpm; slow mixing for 20 min at 40 rpm; and then sedimentation for 60 min. The ferrate dose applied was 0 – 4 mg/L as Fe, and pH of solutions was adjusted by 0.1 M H2SO4 or 0.05 M NaOH to 7.0 – 7.5. All experiments were duplicated. For the real wastewater effluents, the jar test protocol was the same as the above stated but ferrate dose sued was 0, 1, 2, 3, and 5 mg/L as Fe and working pH was pH 6 and 8.

Analytical Methods
As shown in Table 1, three analytical methods were employed for different model test samples. Specifically, for model test solution containing iBu only, solid phase extraction (SPE, Oasis HLB cartridges) + liquid chromatography (LC)-mass spectrometry (MS) was employed, which was conducted in the analytical lab of Bodensee-Wasserversorgung (BWV), Germany. In addition, a simple analytical method, SPE+UV/Vis spectrophotometry, was applied to 100 μg/L iBu model water samples. Moreover, for 10 μg/L CIP and mixed 10 μg/L iBu&CIP water samples, SPE+high performance liquid chromatography (HPLC)-UV was proposed.

Solid phase extraction. The treated model test solution samples were filtered through 0.45 μm cellulose nitrate membrane filters (Milipore) and then enriched by solid phase extraction (SPE). Two types of SPE cartridges were employed in the experiment: Strata-X (200 mg/6 mL and 1 g/20 mL, from Phenomenex) and Oasis HLB (5 mL/60 mg, from Waters USA). Oasis HLB cartridges were only used for those model water samples containing iBu only. Generally, for Strata-X 200 mg/6 mL cartridges, the extraction method was: (1) condition: 4 mL methanol (MeOH); (2) equilibrate: 4 mL deionised water; (3) loading samples: desired amount of model water samples under vacuum at a flow rate of 5 – 10 mL/min; (4) wash: 2 mL 50:50 (v/v) MeOH/H2O; (5) dry: 15 min under gentle nitrogen flow; and (6) elute: 2 x 2 mL 2:49:49 (v/v/v) formic acid/MeOH/ACN. Elutes were either re-constituted to 5 mL for UV/Vis spectrophotometric measurement or dried down to 1 mL for HPLC-UV detection.

On the other hand, iBu only model test solutions were extracted by Oasis HLB cartridges with the use of SPE preparation system Gilson GX-271 ASPEC. The programmed SPE procedure was as follows: (1) condition: 3 mL methanol and 3 mL H2O; (2) loading samples: desired amount of water samples at 10 mL/min; (3) wash: 0.5 mL water; (4) dry: 10 min under a gentle stream of nitrogen gas; and (5) elute: 2 x 2 mL 2:49:49 (v/v/v) formic acid/MeOH/ACN. Elutes were either re-constituted to 1 mL for LC/MS analysis. Prior to extraction, selected samples were spiked with 25 μL of thidiazuron solution (20 ng/μL) as an internal standard.

For the real wastewater treatment, the samples were filtered by 1.2 μm glass fibre filter papers (Fisher), the filtrates were then filtered by 0.45 μm membrane filters (Milipore). The Tandem SPE cartridges used were Strata-X 1 g/12 mL+ENV+500 mg/6 mL, and the extraction procedures were: (1) Adjusted the pH of solution to 2 by 2 M H2SO4, spiked with 1 mL deuterated internal standards; (2) 10 mL methanol +10 mL water were added for the conditioning; (3) flow rate was 10 mL/min for loading samples; (4) 10 mL water was used for washing samples and (5) Drying the cartridges under N2 flow. After extraction, the SPE cartridges were labelled and kept in the freezer for future elution.

The elution was conducted on a Phenomenex SPE 24-position vacuum manifold. The solvents used for the elution are shown in Table 2, and all fractions are collected in silanised vials, combined and dried down at 50°C under a stream of N2.

UV/Vis spectrophotometry. The concentrations of CIP in the 5-mL Strata-X SPE elutes could be measured by a UV/Vis spectrophotometer (Jenway 6505 with 10 mm light-path) at the wavelength of 280 nm. A six-point calibration curve was generated based on absorption of standard CIP solutions in 2:49:49 (v/v/v) formic acid/MeOH/ACN at 280 nm. Concentrations of CIP in model wastewater samples were calculated based on the absorbance reading and the established calibration curve [41].

Table 1. Model test solutions and their specific analytical methods.

<table>
<thead>
<tr>
<th>No</th>
<th>Model water samples</th>
<th>Analytical methods</th>
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<tbody>
<tr>
<td>1</td>
<td>100 μg/L iBu</td>
<td>Oasis HLB SPE+LC-MS</td>
</tr>
<tr>
<td>2</td>
<td>10 μg/L iBu</td>
<td>Oasis HLB SPE+LC-MS</td>
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<tr>
<td>3</td>
<td>100 μg/L CIP</td>
<td>Strata-X SPE+UV/Vis</td>
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<td>4</td>
<td>10 μg/L CIP</td>
<td>Strata-X SPE+UV/Vis</td>
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<tr>
<td>5</td>
<td>10 μg/L iBu+10 μg/L CIP</td>
<td>Strata-X SPE+HPLC-UV</td>
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HPLC-UV analysis. The HPLC separations were performed on the Agilent 1100 system (Agilent Technologies) consisting of a degasser, a binary pump, a thermostated column oven and a diode array detector (DAD). 50 μL of samples was manually injected to a 2.6 μm, 100 mm×2.10 mm reversed phase Kinetex XB-C18 column (Phenomenex). The column was kept at 25°C and eluted with 0.1% formic acid in deionised water and acetonitrile (ACN) at a flow rate of 0.2 mL/min. The elution started with 20% ACN and then with a linear gradient from 20% to 30% ACN over the next 3 min. Then the percentage of ACN was raised to 55% in 2 min, held at this percentage for 9 min and finally lowered to 20% in 1 min. Before the next injection, the system was allowed to equilibrate for 10 min. The DAD wavelengths for IBU and CIP detection were set at 220 nm and 280 nm, respectively.

LC-MS analysis. The LC-MS instrument employed to analyse IBU was Acquity LCT Premier XE system which consists of ACQUITY ultra performance liquid chromatography (UPLC) and orthogonal acceleration time-of-flight (oa-TOF) mass spectrometer (MS) (Waters, USA). The UPLC contained ACQUITY UPLC High Strength Silica (HSS) T3 Column (100 mm×2.1 mm, 1.7 μm particles, Waters, USA). The mobile phase was a mixture of two solvents (Solvent A: 0.1% formic acid in water; solvent B: 0.1% formic acid in ACN). For the analysis of IBU, the elution of the column started from initially 5% B, then a linear gradient increased to 90% B over the course of 7 min, then consistent till 8 min, and finally lowered back to 5% B at 8.1 min. The injection volume of sample was 250 μL, with the running time of 10 min and a flow rate of 0.3 mL/min. After LC separation, the MS analyser for IBU was ESI- mode, with the following parameters: capillary 1823.0 V, sample cone 30.0 V, desolvation temperature 350.0°C, source temp 150.0°C, and MCP detector 2600.0 V. Parent ions (MH+) were monitored at m/z 221.0497.

Results and Discussion
Analytical Methods
HPLC-UV. IBU and CIP were separated with a C18 column. The gradient elution of the column presented a clear separation of both chemicals in 15 min. CIP was eluted firstly from the column with a retention time 9.5 min. The signal of IBU at 220 nm showed a decreasing trend from 5 min and went flat again at 10.5 min. This phenomenon might be caused by the instrumental response to the elution gradient. However, the bulk of IBU came out at around 12.7 min, which was not affected by the previous decreasing signal.

The calibration of IBU and CIP were carried out simultaneously with standard solutions containing IBU and CIP. The curve covered the concentration range of 0.5–6 mg/L, and was conducted four times. The IBU calibration curve with the linear regression is $y = 80.18x + 0.05$, where $x$ means the concentration of IBU and $y$ means the response of instrument, with a coefficient of correlation ($r^2$) 0.997. In terms of CIP, an equation $y = 28.11x - 4.35$ could fit the seven-point calibration well with the coefficient of correlation ($r^2$) 0.990.

Recovery studies of standard solutions ($n = 4$) showed 78.0±14.7% of CIP and 139.1±2.2% of IBU recovered by the instrument, respectively.

LC-MS. A seven-point calibration curve for the concentration range of 0.04–1.2 mg/L was generated based on MS response of standard IBU solutions in methanol. Quantification of IBU was based on the response at m/z 221.0497 (MH+) vs. linear regression equation $y = 57.76x$, where $x$ means the concentration of IBU and $y$ means the response of the instrument, with the coefficient of correlation 0.998. Recovery was obtained by running standard IBU solutions (5 μg/L) through SPE and LC-MS process, with the mean recovery 117.7±30.6% ($n = 4$).

Single Compound with Initial Concentration 100 μg/L
For CIP samples with initial concentration of 100 μg/L, average 60% of CIP was removed from the model wastewater with a low ferrate (VI) dose ranging from 0.02 to 0.34 mg/L as Fe (Fig. 1). As increasing the dose of ferrate (VI), slight improvement in the removal efficiency was observed, from 61.2% at 0.02 mg Fe/L to 68.9% at 0.34 mg Fe/L. The average removal efficiency dropped dramatically to 53.0% at dose of 0.25 mg/L as Fe may be caused by the analytical deviation. Though the overall removal of CIP at this dose range was less than 70%, the benefits of using ferrate were apparent in that low ferrate doses (maximum 0.34 mg/L as Fe) can remove up to 69% of CIP.

Further studies were carried out to investigate if relatively high ferrate doses could improve CIP removal. Figures 1 and 2 show that over 80% of CIP can be removed if the ferrate dose reached or exceeded 1 mg/L as Fe. This is consistent with that from a preliminary study [42] where a small dose of ferrate (up to 1 mg/L as Fe) can remove more than 80% of CIP and residual concentration could be lower than 1 μg/L.

However, the treating performance of IBU by ferrate (VI) was not as good as that of CIP, less than 25% of IBU was removed from the model water when dosed ferrate (VI) was 0.5–4 mg Fe/L (Fig. 2). The highest removal of IBU happened at the dosage 3 mg/L as Fe, where 21.0±4.9% of IBU was degraded.

Single Compound with Initial Concentration 10 μg/L
Compared with the case of 100 μg/L, CIP model test solutions, when the initial concentration was 10 μg/L, the overall treating performance of CIP by ferrate (VI) decreased to less than 60% (Fig. 3). Specifically, when the ferrate (VI) doses were 0.5–2.5 mg Fe/L, the CIP removal efficiencies were between 25% and 60%, with a highest removal 55.5±1.2% at the dose of 2.0 mg/L as Fe.

On the other hand, the overall treating performance of IBU improved when the initial concentration was 10 μg/L. As shown in Fig. 3, the IBU removal at ferrate dose 0.5 mg Fe/L was 11.2%, which was similar to the situation when the initial IBU was 100 μg/L. Raising ferrate dose improved the IBU removal. When the dose was over 1.5 mg/L as Fe, the IBU removal could exceed 25%, with the maximum IBU removal of 40% for an optimum dose, 1.5 mg Fe/L.

Mixed IBU&CIP
Further study was carried out to investigate the treating performance of mixed IBU and CIP model test solutions by ferrate. The initial concentrations of both compounds were 10 μg/L, and the ferrate doses applied were 0.5–2.5 mg/L. Results show that ferrate salts possess more capability to remove CIP than IBU (Fig. 4). Averagely, at each dose, ferrate removed CIP around 10% more than IBU. The overall removal of IBU was less than 15%,
Figure 1. CIP removal by low doses and relatively high doses of ferrate (VI).

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Figure 2. IBU and CIP removal by ferrate (VI) doses of 0.5–4 mg/L as Fe.

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with the highest removal efficiency 15.2±1.8% at dose 2.0 mg/L as Fe. The treatment of CIP was improved by increasing the ferrate dose; when the dose increased to 2.5 mg Fe/L, the CIP removal increased to 31.1±6.6%.

Fig. 5 gives the comparison of the pharmaceutical removal among different model test solutions. Generally, the treating performance of samples with single compound was better than those with both compounds. For instance, at each dose, the CIP removal of samples with single compound was 7–30% higher than that of samples with both compounds. The biggest gap happened at dose 2.0 mg/L, where CIP removal for CIP only samples was 55.5±1.2% while for mixed samples was only 25.9±3.4%. In the case of IBU, the IBU removal in the IBU alone solutions was 5–35% higher than that of in the mixing IBU and CIP solutions.

Ferrate (VI) has shown much more capability in the degradation of CIP than that of IBU. This can be explained that CIP belongs to electron-rich organic moieties (EOM) which can be easily transformed during ferrate oxidation. However, the carboxylic group in IBU is an electron-withdrawing group with which the reactivity of ferrate is usually slow [35,43].

**pH Dependence**

Preliminary studies showed that comparing with initial pH, final solution pH after dosing ferrate has much more influence on the removal of CIP and IBU. As shown in Fig. 6, IBU removal showed great difference between pH 4 and pH 10. At pH 4, for all three doses of ferrate, IBU removal was much higher than other pH conditions, with over 50% IBU removal for the dose of 1 mg/L or 2 mg/L. However, for nearly all samples with final pH over 4, IBU removal was below 30%. As shown in Figure 7, for all ferrate doses, the CIP removal efficiencies of samples with final pH between 4 and 8 were above 45% and were much higher than those of final pH 9 and 10 (less than 25% CIP removal).

The effect of final solution pH on the IBU or CIP removal might be understood by considering compounds pH values. The pKa of IBU is 4.91, therefore when the pH was <8, the major species would be IBU, which was relatively easy to be oxidised. For CIP, its pKa is 6.09 and 8.2, respectively, and when the pH of solution was >8, the major species of CIP would be dissociated CIP-O\(^{-}\), which was relatively difficult to be oxidised [38], and this leads to an overall decrease in CIP removal efficiency. Strong pH dependence of removal efficiency of IBU and CIP was also observed by other researchers. The reaction rate constant of ferrate (VI) versus ibuprofen (IBU) was 0.3 M\(^{-1}\) s\(^{-1}\) at pH 7.5 then slipped sharply to 0.01 M\(^{-1}\) s\(^{-1}\) at pH 9.0 [43], and the rate constant of ferrate vs. ciprofloxacin (CIP) was over 500 M\(^{-1}\) s\(^{-1}\) at pH 6 but only 10 M\(^{-1}\) s\(^{-1}\) at pH 9.5 [35]. In general, both IBU and CIP belong to electron-rich organic moieties (EOM), which can be potentially transformed during ferrate oxidation and this has been demonstrated in other studies (e.g., [44]).

Strong pH effect on the IBU and CIP removal by ferrate(VI) can also be considered from ferrate(VI) speciation against solution pH. There are four ferrate(VI) species in aqueous solution that depend on pH: H\(_2\)FeO\(_4\)\(^+\), H\(_3\)FeO\(_4\), HFeO\(_4\)\(^-\), and FeO\(_4\)\(^{2-}\), and the corresponding dissociation constants are pK\(_1\) 1.6±0.2, pK\(_2\) 3.5, and pK\(_3\) 7.23, respectively [45]. FeO\(_4\)\(^{2-}\) is the dominant species in alkaline conditions, and HFeO\(_4\)\(^-\) predominates in mildly acidic conditions. Ferrate(VI) has a higher oxidation potential at low pH (2.2 V) than in the alkaline condition (0.72 V) [23] and thus the lower the solution final pH, the stronger oxidation potential of ferrate(VI) [HFeO\(_4\)\(^-\)] predominates although the stability of ferrate(VI) decreases at low pH solutions.

![Figure 3. Treatment of 10 \(\mu\)g/L IBU model water samples by ferrate (VI) (pH = 7.05±0.25).](https://doi.org/10.1371/journal.pone.0055729.g003)
Figure 4. Experimental results of 10 μg/L IBU and CIP mixed model test solutions (pH = 7.05 ± 0.25).
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Figure 5. Comparison of the treating performance among different model test solutions (pH = 7.05 ± 0.25).
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Figure 6. pH dependence of IBU removal by ferrate.
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Figure 7. CIP removal of samples with various final pH for various ferrate doses.
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Removal of Selected Pharmaceuticals Spiked in the Final Effluent by Ferrate (VI)

Occurrence of pharmaceuticals in the raw effluent. Six compounds were detected in the raw effluent samples, with concentrations ranging from 100 to 320 ng/L. The concentrations of naproxen, carbamazepine and atenolol were over 200 ng/L, among which naproxen exhibited the highest concentration with an average concentration 317.3 ng/L. In addition, erythromycin-H2O, lidocaine and bezafibrate were also detected in the effluent, with the concentrations 100–120 ng/L.

Removal of pharmaceuticals at pH 6 and 8. As shown in Fig. 8, except ciprofloxacin and naproxen, the removal efficiencies for other compounds by ferrate were under 25% in the dosage range when the solution pH was 6. As for ciprofloxacin, the removal efficiency showed good linear correlation with the rising ferrate dose, e.g. from 16% at 1 mg/L as Fe to 69% at 5 mg/L as Fe.

Removal efficiencies of ciprofloxacin, naproxen and ibuprofen were 20–60% for ferrate doses 1–5 mg Fe/L. However, the elimination of n-acetyl sulphamethoxazole was only less than 10% for any ferrate dose. Besides, except ciprofloxacin and ifosfamide, raising ferrate dose did not improve the pharmaceutical removal significantly. As for ifosfamide and ciprofloxacin, when the ferrate dose was raised from 3 mg/L to 5 mg/L, the removal efficiency increased from 9% to 38% for ifosfamide and from 44% to 63% for ciprofloxacin, respectively (Fig. 9).

Fig. 10 shows the pharmaceuticals removal by ferrate when the solution pH was 8 and ferrate dose was 5 mg/L as Fe. As shown in the graph, the removal efficiency of ciprofloxacin was the highest among the compounds listed, with an average removal of 63%, followed by naproxen (43%). On the other hand, n-acetyl sulphamethoxazole was the hardest to remove by ferrate, with an average removal of 8%. The selective removal of pharmaceuticals by ferrate(VI), as demonstrated by this study, is consistent with that from other studies [35]. Ferrate(VI) can degrade electron-rich organic moieties [35,39] of pharmaceuticals such as ciprofloxacin and then achieve high percentage removals but difficult to oxidise other pharmaceuticals such as n-acetyl sulphamethoxazole and ibuprofen. Nevertheless, sewage tertiary treatment by ferrate(VI) should achieve the removal of pharmaceuticals and other emerging micro pollutants as well as will enhance the effluent qualities in general such as lower concentrations of suspended solids, phosphate and COD [30].

Conclusions

Results of treatment of CIP by ferrate show that ferrate can remove at least 60% of CIP from model test solutions even at very low ferrate doses (<0.3 mg/L). Besides, increasing ferrate dose up to 1 mg Fe/L as Fe can achieve more than 80% removal efficiency of CIP. However, ferrate did not show similar treating performance for IBU degradation, with only 30% IBU removal at ferrate dose of 2 mg Fe/L.

The final solution pH affects the treating performance significantly for both pharmaceuticals. When final pH of solution was greater than 8, CIP removal efficiency by ferrate drops significantly. In the case of IBU, pH 4 was the optimum pH for IBU removal by ferrate.

Ferrate has shown higher capability in the degradation of CIP than IBU. This is due to that CIP belongs to electron-rich organic moieties (EOM) which can be readily degraded by ferrate oxidation. Instead, the reactivity of ferrate with electron-withdrawing groups (carboxylic groups) in IBU is slow and consequently, lowers ferrate removal efficiency for the IBU.

Ferrate has shown promising performance at both pH 6 and 8 and dose range of 1–5 mg Fe/L in the treatment of real waste.
Figure 9. Pharmaceuticals removal by ferrate at pH 8.
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Figure 10. Pharmaceuticals removal by ferrate at pH 8 and ferrate dose 5 mg/L as Fe.
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water effluent. Removal efficiency of ciprofloxacin was the highest among the target compounds, with an average removal of 63%, followed by naproxen (43%). On the other hand, n-acetyl sulphamethoxazole was the hardest to remove by ferrate, with an average removal of 8%. Furthermore, the removal efficiencies of other compounds by ferrate were between 20% and 40%.

References


Author Contributions

Conceived and designed the experiments: JQJ ZZ. Performed the experiments: ZZ JQJ. Analyzed the data: JQJ ZZ. Contributed reagents/materials/analysis tools: JQJ ZZ. Wrote the paper: JQJ ZZ.